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Synthesis and absolute stereochemistry of spiroacetals in rove beetles (*Coleoptera: Staphylinidae*)

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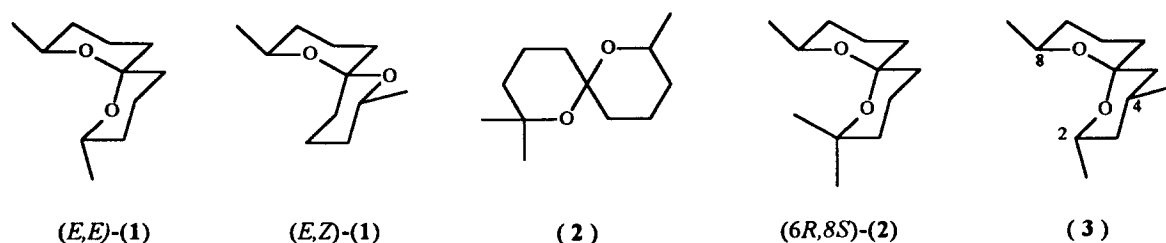
Abstract

The unusual branched-carbon chain spiroacetal, 2,2,8-trimethyl-1,7-dioxaspiro[5.5]undecane, has been synthesised as its racemate and (6*S*,8*R*)-isomer. The natural compound, identified in the rove beetle, *Ontholestes murinus* (L.) proved to be the (6*R*,8*S*)-isomer. (*E,E*)-2,8-Dimethyl-1,7-dioxaspiro[5.5]undecane, a major component of the volatiles from the same insect, is the (2*S*,6*R*,8*S*)-isomer, but is largely racemic in *Ontholestes tessellatus* (Geoffr. Fourcr.). © 1999 Elsevier Science Ltd. All rights reserved.

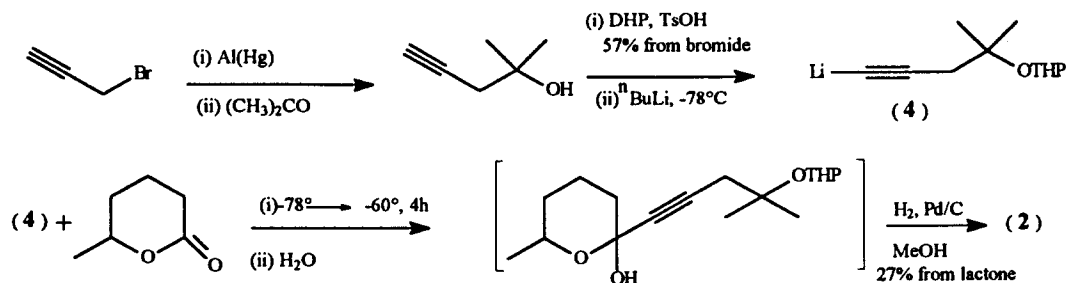
Keywords: insect; spiroacetal; enantioselective chromatography.

The defensive strategy evolved by rove beetles (*Coleoptera: Staphylinidae*) involves abdominal elevation and the discharge of a secretion from a pair of everted glands, in the direction of the molestation. In 1986, Dettner and Schwinger¹ described the defensive secretion of *Ontholestes murinus* (L.), which contained (*E,E*)-2,8-dimethyl-1,7-dioxaspiro[5.5]undecane (M=184) (**1**) of undetermined stereochemistry. This spiroacetal was accompanied by another component with an apparent molecular mass of 198, and a spiroacetal-like mass spectral fragmentation pattern.² In a subsequent report,³ this latter compound was described as a 2,2,8-trimethyl-1,7-dioxaspiro[5.5]undecane (**2**) without further data. In view of the fact that branched-carbon chain spiroacetals are rare in insects^{4,5} (only two are now known),⁶ we have undertaken syntheses which permit the determination of the absolute configuration of this unusual compound **2**, as well as that of **1**. The other insect-derived, branched-carbon chain spiroacetal is (2*S*,4*R*,6*R*,8*S*)-2,4,8-trimethyl-1,7-dioxaspiro[5.5]undecane (**3**), the major component of the abdominal gland secretion of the shield bug, *Cantao parentum* (White).^{7,8}

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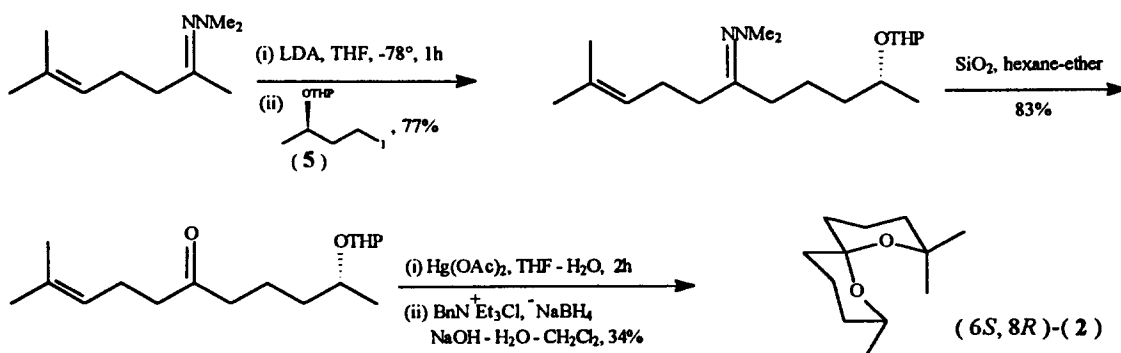
Racemic **2** was synthesised in a straightforward way by addition of the lithium salt of the protected ynol (**4**), to δ -caprolactone, followed by hydrogenation–deprotection and cyclisation⁹ of the initial addition product, in 27% overall yield from the lactone (Scheme 1).



Scheme 1.

Final purification of **2**¹⁰ was achieved by flash chromatography on silica gel (30:1, hexane:ether) followed by preparative gas chromatography. The enantiomers of **2** were very well separated on a J&W Cyclodex-B Column (conditions: splitless injection, 40°C for 2 min then 20°C/min to 120°C then 1°C/min).

Acquisition of the $(6S,8R)$ -enantiomer of **2** was based on hydrazone alkylation with the (R) -iodide (**5**),¹¹ followed by an oxymercuration–deprotection–cyclisation sequence¹² as shown below (Scheme 2). The resulting $(6S,8R)$ -enantiomer ($[\alpha]_D^{22} +46.2$, c 7.09, CHCl_3),¹³ thus produced, exhibited MS and NMR parameters identical with those of (racemic) **2**, and eluted earlier than its antipode. The EI mass spectrum of synthesised $(6S,8R)$ -**2** is shown in Fig. 1.



Scheme 2.

Three gland reservoirs from *O. murinus* were crushed in hexane and examined by high quality GC–MS. A low level of 2,2,8-trimethyl-1,7-dioxaspiro[5.5]undecane (**2**) was identified, and comparisons with synthesised racemic **2** and $(6S,8R)$ -**2** demonstrated that the natural **2** was the $(6R,8S)$ -isomer of >95% ee. The accompanying (E,E) -**1** was the $(2S,6R,8S)$ -enantiomer (as drawn above) of >98% ee.

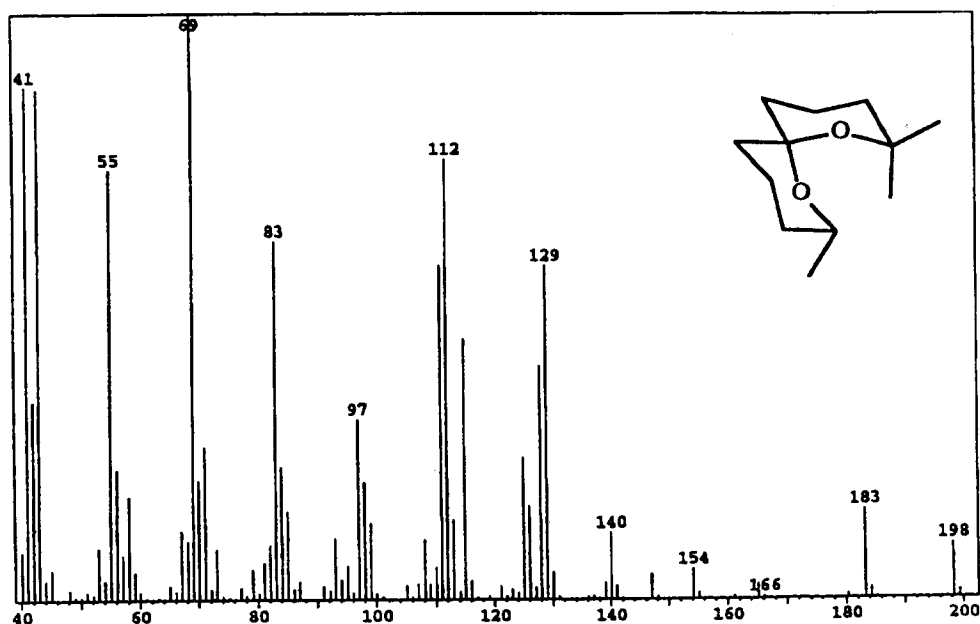


Figure 1. EI mass spectrum of 2,2,8-trimethyl-1,7-dioxaspiro[5.5]undecane (2)

Another rove beetle, *O. tessellatus*, also contains (*E,E*)-2,8-dimethyl-1,7-dioxaspiro[5.5]undecane (1), and GC-MS comparisons (J&W Cylodex-B, as before) with authentic samples revealed comparable levels of the widely occurring (*2S,6R,8S*)-isomer and its (*2R,6S,8R*) antipode, with the latter unreported from an insect source.^{5,6} In some respects, the presence of the (*2R,6S,8R*)-isomer is not too surprising, as (*E,Z*)-1, which often accompanies^{5,6} (*E,E*)-1, has opposite configurations at carbon atoms 2 and 8, confirming the possibility of both (*R*)- and (*S*)-hydroxylation modes. This outcome can be accommodated within the hypothesis recently proposed¹⁴ for spiroacetal biosynthesis in fruit-fly species, if fundamentally similar processes are operating. The defensive secretions of rove beetles of the sub-tribe Staphylinina are dominated by biologically active iridoid compounds,³ but the biological significance of the spiroacetals in these defensive glands or secretions is not known. They are inactive if applied topically but exhibit a drastic insecticidal fumigancy at very low vapour concentrations towards adults of *Drosophila melanogaster*.¹⁵ It is remarkable how diverse are the components of these defensive secretions and also the structure and stereochemistry of the spiroacetals utilised. Spiroacetal (2) co-occurs in *O. murinus* with terpenoid components, and its structure in part may be isoprenoid derived, however, biosynthesis involving valine or leucine may also be possible.

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2. Francke, W.; Hindorf, G.; Reith, W. *Naturwissenschaften* **1979**, *66*, 618. Comparison with the mass spectra of known spiroacetals with M=198 showed no coincidence. A notable feature of the mass spectrum of the unknown compound was the relatively high abundance of the M-15 ion (*m/z* 183), when compared with the M-15 ion for 1. This indicated an especially stabilised oxonium ion, possibly arising from methyl loss from a *gem*-dimethyl group adjacent to oxygen. For a plotted 70 ev mass spectrum of (*E,E*)-1, see: Francke, W.; Reith, W.; Bergström, G.; Tengö, J. *Naturwissenschaften* **1980**, *67*, 619.
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10. Compound **2**: For EIMS, see Fig. 1. HRMS: $C_{12}H_{22}O_2$ requires: 198.1620. Measured: 198.1616. 1H NMR (200 MHz, $CDCl_3$): δ 1.04 (d, $J=6.3$, 3H), 1.13 (s, 3H), 1.30 (s, 3H), 1.32–2.02 (m, 12H), 3.82–3.92 (m, 1H). ^{13}C NMR (50 MHz, $CDCl_3$) δ : 15.8, 19.2, 21.5, 25.3, 32.8, 32.9, 36.4, 37.1, 37.2, 65.7, 72.0, 96.2.
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